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Australasian Mycological Newsletter

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SOME PRACTICAL SUGGESTIONS FOR FUNGAL STUDIES

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These procedures have been accumulated over a number of years—mostly from trial and error (large amounts of error it always seems in retrospect) when working either in the field or with either fresh or dried material under the microscope. Admittedly, since my field of study is the agarics, the suggestions here are derived from those studies, but they can generally be easily extrapolated to any mycologist's work. Some, if not all of these ideas will be obvious to and already being used by the workers who have been dealing with the fungi for many years and already I can hear the statement: 'I know that—it's an oldie'. But I ask those people to bear with me; I can only say that once upon a time it wasn't an 'oldie' to them either and all of us forget the frustration we often faced as we struggled to obtain results when everything we tried to do seemed to go wrong.

The problem is that for people just starting to traverse the 'minefields of mycology', there is no 'compendium' of tricks of the trade that can be appealed to when things just *don't* go right and nothing that is tried seems to help. Sooner or later, something is done which allows the work to proceed—but the method is then stored in the mind of the worker and never committed to print—or at best disseminated by word of mouth. This little script tries to collate a few tips. Not all of the tricks will be covered and if the reader knows of others, please let me know so that for a later edition they can be included.

Field work

Dealing with those 'Very Slight Odour' Fungi

Smells are notoriously difficult to describe because everyone's sense of smell is different: I love the smell of jasmine for example, but my wife detests it and even finds the odour painful to breathe. Fungi are often described as having various smells, including a 'fungous' odour that we all recognise but find it difficult to put in words. Some fungi are described as having an odour but in the field the worker often cannot detect it. A classic case is that of *Cantharellus concinnus* (= *C. cibarius* var. *australiensis*). This definitely does have a distinct apricot smell that is immediately detectable by some (including myself) but others struggle in vain to notice anything at all. A very good way of enhancing the odour is to put the collection in a very small, clean plastic container for half an hour, then carefully open the lid a fraction and sniff the air at the opening. Any odour is concentrated and will be rapidly detected. This has proven an excellent method of proving the apricot odour of the above species even to 'non-believers'.

Detecting Viscidity of the Pileus and/or Stipe Using the 'Lip Test'

Many species are obviously viscid or sticky with a layer of gluten over the pileus and stipe, or they have gelatinised surface hyphae which also create this state. Unfortunately, many species rapidly lose this viscosity as they mature, sometimes because the humid conditions that were present at first have vanished and the pileus and stipe are now dry and sometimes because that is how the particular species develops. Often the pilei of these species have a varnished appearance that gives them away, but equally as often there is nothing to suggest the original viscosity. This becomes very irritating as many keys quite happily ask the user to choose whether the pileus (or stipe) was viscid and if there is no way to tell if the viscosity was originally there, the key user comes to a full stop.

Most people use their finger to test fresh material for the stickiness of the viscid pileus. However, for these difficult species, the finger is not sensitive enough. The best method to detect, in the field, the presence of a viscid cuticle is to use the 'lip test'. Touch the fungus gently onto the very sensitive skin of the upper lip surface and then equally gently and slowly pull it away—if there is any sticking sensation it will be very easily detected and a viscid pileipellis can be recorded. If the cutis is dry, a small drop of distilled water (never use saliva as this has enzymes in it that may alter the pileal surface) will revive the viscosity sufficiently. In all the cases so far encountered by the author, the 'lip test' has never failed to confirm a suspected ixocutis or ixotrichoderm.

(My apologies to the person who told me of this technique—I cannot remember who you are but thanks very much for the information.)

Getting that Fungus out in the Field

There are various suggestions that are made as regards how you should equip yourself in the field. I have tried numerous methods, but I would like to share the tried and true ones that I now seem to use very consistently. As regards collecting the fungus, most authorities recommend carefully digging it up with a tiny bit of soil if it is a ground species. This is so you can see if there is a volva that is critically important for the genus *Amanita* or any other species with below ground structures. You can carry a small garden trowel or a specially made implement, but my preference is now an old and battered 'Swiss army penknife', carried in a belt pouch. Before anyone asks, it has only a single, rather battered blade and nothing else—I've yet to see the fungus that needs to be extracted with a corkscrew. This knife blade allows all the digging normally required. It can remove species from wood (except of course, those like *Phellinus* which need a small axe) and it has the added advantage that when you want to have lunch, it can be cleaned and used to butter your sandwich and peel the apple or orange. Even very deep fungi can be removed using this

small blade. The most spectacular case was a *Cordyceps gunnii* which needed about 30 cm of painstaking and very careful excavation through a tree fern trunk and then into the soil.

A small, 10–20 hand lens is essential. It allows examination of quite small fungi in order to see details that may sometimes be absolutely critical in the field before the fungus wilts in any way.

I have also found that a set of forceps is absolutely essential. These are used to collect very tiny and delicate species. Forceps are also one of the easiest things to lose and I have not lost my pair of forceps in 10 years. (With one exception that I am sure Alan Mills is delightedly waiting to mention.) What I have done is drilled a hole at the top of the forceps and threaded through this a very brightly coloured cord—orange or yellow for preference as it stands out like a beacon on the forest litter. You just can't lose them or put them down because a final 'look around' always shows the bright cord and the attached forceps. The only time I have ever 'lost' my forceps was when I was in Tasmania in April/May 1998: before my final packing, I emptied a box with some forest litter outside on the ground—the forceps were of course, underneath the litter. Alan found my forceps about two weeks later in the back yard, solely because the bright cord was attached—I have been suitably grateful as I value those particular forceps rather highly. Incidentally, I also use a small piece of plastic tubing placed over the sharp ends of the forceps so that when they are placed back in my top pocket I don't get impaled on the sharp ends.

Storing the fungus to bring it back as carefully as possible is very dependent on how much you want to carry and exactly what it is that you are looking for. If 'anything is grist to your mill', the best things are probably small plastic containers (margarine containers with lids are excellent) and some grease proof paper to wrap up the larger items. If, as in my case, the target fungi are all smallish and delicate, then a very useful container is the 'bait box' with its compartments which can be stacked on the floor of a basket and carried without banging or damaging the specimens.

In the Laboratory

Detecting the Presence of an *Ixocutis* or an *Ixotrichoderm* on Herbarium Material

One of the most aggravating things is trying to decide if the 'withered piece of agaric fungus' being studied, originally had a viscid cuticle on the pileus. A very good starting procedure is always, and I **do** mean *always*, examine the surface of the pileus using a stereo dissecting microscope under about 20–40 magnification. From considerable experience, an originally viscid pileipellis will almost always make its presence known by a 'varnished wrinkled' appearance under this magnification. The surface looks 'polished' as a result of the dried 'gluten' (or whatever caused the viscosity) and as the surface dries, it produces a very distinctive 'skin' over the surface that wrinkles. Over perhaps 300–400 collections examined this way, I am unable to re-call any collections that demonstrated this character which did **not** eventually prove to have had a viscid pileus when first forming in the field.

Cutting a Section

I suspect that most agaricologists find that hand sections are quite good enough for what has to be examined as regards sections of pilei or lamellae. It takes practice and a stereo dissecting microscope, but once mastered the technique is simple and extremely effective. I never bother with scalpel blades—unless there is *absolutely* nothing else—because I have found they are too thick. Better blades can always be obtained from the old, 'double edged' razor blades such as Gillette or Wilkinsons. The original Gillette blades were made of high carbon steel and snapped beautifully, however, the modern stainless steel blades will not do this very successfully. I find the best way is to have a very small pair of scissors devoted to 'tin-snip' work and cut very small, scalpel shaped sections of a blade as I require them. These tiny pieces of blade can then be mounted in the handles of dissecting needle holders. Use the screw-ended ones and then you can replace the used blade sections as often as you need to.

Recently I also found another source of even better blades. The double bladed 'plastic combinations' that are used in modern pivoting razors have an even finer metal foil than the old Gillette type twin blades. I very carefully pulled apart a blade after considerable bathroom use and even then found that the edges on these used blades were superior for cutting very fine sections when compared with new twin blades. My thanks to Katrina Syme who first informed me of this method in July 1998.

Tweezers and Dissecting Needles

No matter how you try, needle-pointed tweezers still end up looking like crowbars under a stereo microscope. This is all the more irritating if you have a very fine section just cut and the result is rather like 'trying to delicately manipulate a postage stamp-sized piece of tissue paper with a pair of fencing pliers'. One way out of this is to use the razor blades that have been cut with your dedicated 'tin-snip scissors'. You can cut very tiny pieces of blade and due to the action of the scissors, these tiny pieces are also a little curled. Mount one of these in a dissecting needle holder. The result is a very fine, almost hook like manipulator which can be used to tease out the hand cut section and very delicately place it on the microscope slide.

Rehydrating a Section While Keeping its Side View Position

When you are just rehydrating a 'lump of tissue' and only intend to splay it out, then there is no need to worry about placement of the section on the slide. However, if the material to be examined is a carefully cut hand section and it is critical that the details of the section be seen from the side, then just placing it on the slide is absolutely out of the question. From very, very bitter experience, the traditional method of 'place the section on the slide, add a drop of reconstituting mountant—such as ammoniated congo red—and then place a cover slip delicately on top' is just not good enough. If you do use this method, you will find it will work successfully in about 30–40 per cent of the attempts; the remainder of the time, the material twists as soon as the mountant strikes it and when the cover slip is put on, the twisted material has taken up a position which usually negates the effort put into getting as thin a section as possible and you have to start all over again—often thinking some very interesting words about mycology in general.

A large number of very frustrating moments can be avoided by the most simple and obvious procedure—but again, it's so obvious that no one ever tells you. Once the section is cut, place it on the microscope slide. Now, put the cover slip on the dry material and check to see that it is still flat—use the stereo microscope. Next, place a drop of the reconstituting mountant on the slide at the edge of the cover slip. It will immediately be drawn under the cover slip and rehydrate the section. Because the cover slip is present to hold the section firmly, it cannot twist or distort and virtually a 100 per cent success rate is guaranteed using this method of looking at sections.

Keys, Keys and 'Schinken' Keys

I remember once about 15–20 years ago, when I was ever so naive, that I thought a single key was all I needed to completely discover the world of fungi. Better yet, I thought that a key—any key—was absolutely correct in all details. Ummmmmm...how things change. About 10 years ago, I recall spending an afternoon in my study with Alec Wood while we pored for about two hours over a collection obtained that morning from the Bunya Mountains. We were using the agaric keys published by a very well known and world respected agaricologist—who shall remain nameless. We believed that what we had was a species of the genus *Marasmiellus* because all the characters, apart from one, were present. But after examining the pileus' cuticle something like five or six times we could find nothing of the rameales hyphal structure that was supposed to be present. The problem was that our main key said that the hyphae had to be there—or it wasn't in the genus. After two hours of brick walls, we mentally 'tore our hair' and decided that it was a *Marasmiellus* whether the key to the genera agreed or not. So we finally went to the sub-section that contained the genus *Marasmiellus* and there, part way down the key to the species was a couplet which essentially said: 'Rameales hyphae absent from the pileus in this small group'. I forget exactly what we said following this discovery but it meant that although the group was known to the mycologist who constructed the keys, there was no way of getting to this group validly from the main part of the keys.

The lesson was re-learned fairly abruptly. Don't just use one key on a new species, try several by different authors and if about four out of the six give the same answer, you are probably on to the correct result. If you come to a dead end, try some different paths and assumptions because quite often these will eventually lead to the correct answer.

'Photographing the Brute'

Most of my photographs are taken in the laboratory under contrived conditions. This is because mostly I only require photographs that allow diagnostic processes, not 'prettiness' of the subject. Nevertheless, there is a place for these pretty photos and in any event, without them, the fragile beauty of these natural gems would not be appreciated by the larger public—something to be greatly desired and encouraged. However, a couple of tips in the photographic area are worth knowing, especially if they can save money.

I mostly use colour slide film because I can show the pictures on screens for audiences, I can always get colour photographs made, and I can scan the slides easily into a computer. If possible, the film I use is Kodachrome 25ASA. This is extremely slow but very, very fine-grained and therefore allows a scanner to produce excellent images from the slide. Of course, this means that most field photographs must be taken by flash. If 25ASA is unobtainable (it is still made and can be got in specially by any good camera store if they normally don't stock it) then 64ASA is almost as good.

The ultimate way of photographing fungi in the laboratory is to use a copying stand. This allows the camera to be positioned directly above the specimen(s) and held absolutely still. The specimens can be arranged nicely and the picture taken with a flash attachment. A copying stand is quite an expensive item, between \$300 and \$800, but a simple multi-purpose copying stand can be had for about \$50. What you do is purchase a tripod but make sure that it is one of the type that will allow you to take the central adjusting column out and invert it *i.e.* suspend the camera from

underneath and inside the tripod legs. The specimen is now set up inside the tripod legs and the camera attached. There will be a few 'bugs' that have to be worked out of the system—including how to see down through the view-finder, and of course making sure that the camera does not fall downwards under any circumstances—but they are not insurmountable. Always use a shutter release cable to take the photograph and as a general rule, use a flash diffuser over the flash bulbs to soften the light and reduce 'blooming' where large glows of light appear on the photographic slide. This method was the one I used extremely successfully throughout my Tasmanian trip during 1998 and the results were all I could have wished for.

Some extra tips on the photography include the fact that the fungi should be placed on a ground glass sheet about 20 cm off the ground so that shadows are not photographed. The background should be a neutral grey colour. If you use a photographer's standard grey, the slides you take can produce images that can always be brought back to the original colour.

Drying the material for preservation

I have found two methods which are equally suitable depending upon where you are situated. My home is solar powered and normally I don't like using electrical heating elements to dry fungal specimens—a heating element is one of the most energy consuming methods of employing electricity. Instead I use a gas-powered dryer made of freshwater tank strainers fitted with a pilot light from a gas fired hot water system. The pilot light is also fitted with a small hot plate that acts as a heat diffuser. My specimens are usually dry overnight and the amount of gas used is extremely small: a one and a half kilogram gas bottle is good for about a week's continuous drying use.

The other method was introduced to me when I was at the Wilson's Promontory meeting of the Victorian Field Naturalist's group in May. I had never thought of this one but seeing the results has resulted in my now having my own drier which is taken with me if I am ever collecting in a situation where I have access to mains electricity. The drier is simply a proprietary brand food drier. These have a series of temperature settings from about 35–50 degrees and blow the warmed air over the fungal material that is placed on a series of racks. These driers are very effective and contrary to my misgivings, the constant air flow doesn't seem to mix the spores from any of the fungi to any extent. I have to thank Tom May for introducing me to that obvious use of a commercially available unit. The cost is (was) under \$150 but I have seen dedicated mycological driers at costs of over \$250.

And finally, again my plea—if you have any good ideas for any area of fungal research—tips that allow better applications of well known techniques, let me know for the future. My email address is generally examined daily if you wish to contact me rapidly.

SOME PRELIMINARY RESULTS FROM COLLECTIONS OF HYGROPHORACEAE FOR THE SEASON 1998

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During the interval April to June 1998, collections of species within the Hygrophoraceae were made in Tasmania, Victoria and New South Wales. Additionally, examination was made of a small number of collections from the Victorian State Herbarium (MEL). Some very preliminary results are now available and species lists recording the occurrence of a particular taxon in one or more of the State locations are given below. Except in special circumstances (e.g. where a taxon has previously not been described or listed for Australia), author citations are assumed to be those as for the taxa contained in Young & Wood (1997):

Tasmania (97 collections)

Hygrocybe anomala, *H. astatogala*, *H. aurantiopallens*, *H. batesii*, *H. cantharellus*, *H. chromolimonea*, *H. erythrocrenata*, *H. firma*, *H. graminicolor*, *H. irrigata* (Pers.: Fr.) Bon (new record for Australia), *H. julietae* (Stevenson) Horak (new record for Australia, still to be confirmed), *H. lewellinae*, *H. lilaceolamellata*, *H. mavis*, *H. miniata*, *H. pseudograminicolor*, *H. reesiae*, *H. rodwayi*, *Hygrophorus involutus*.

There are also approximately 18 new taxa to be described.

Victoria (40 collections)

Hygrocybe astatogala, *H. aurantiopallens*, *H. batesii*, *H. chromolimonea*, *H. conica*, *H. firma*, *H. hayi*, *H. leucogloea*, *H. lewellinae*, *H. mavis*, *H. miniata*, *H. rodwayi*.

There are also approximately 3 new taxa to be described.

New South Wales (130 collections)

Hygrocybe astatogala, *H. aurantiopallens*, *H. aurantipes*, *H. batesii*, *H. cantharellus*, *H. cerasinomutata*, *H. chromolimonea*, *H. erythrocala*, *H. graminicolor*, *H. kula*, *H. lewellinae*, *H. lilaceolamellata*, *H. miniata*, *H. pseudograminicolor*, *H. reesiae*, *H. rodwayi*, *H. sanguineocrenulata*, *H. siccitatopapillata*, *H. stevensoniae*, *H. sylvaria*, *H. virginea*, *Hygrophorus involutus*.

There are also approximately 12 new taxa to be described.

RESULTS

Several interesting results are already apparent from these preliminary details. First it was extremely gratifying to find collections that could be assigned to taxa described in the 1997 paper for which only the holotype collection was known. Amongst these are *Hygrocybe siccitatopapillata*, *H. pseudograminicolor*, *H. sylvaria*, *H. hayi* and *H. leucogloea*. Even more useful was the fact that photographic material could at last be stored for some of these species.

A second result is that the distributions and abundance of some of the taxa can for the first time be assessed, although for many species, there are still too few collections. *Hygrocybe pseudograminicolor* was previously known only from its holotype locality of Mt Wilson in the Blue Mountains west of Sydney. It is now known to be widespread and abundant in Tasmania. Another surprise has been *Hygrocybe lewellinae* which for many years has been considered to be one of our rarer fungi. The species is widespread and abundant in Tasmania and is also very widespread and surprisingly abundant on the mainland.

A third and very curious habit of the Hygrophoraceae was to show that a 'cherished concept' may need to be discarded. The collection plans for 1998 were based around the idea of visiting Tasmania in April–May and collecting there before the season became too cold. The aim was then to cross over to the mainland and 'follow the season northwards' as the cold weather also moved northwards with the approach of mid-winter—assuming of course that normal rains fell. This proved to be an incorrect prediction for fruiting behaviour in this family because the present indications are that once the Hygrophoraceae fruiting season starts in Tasmania, it starts simultaneously all up and down the eastern coastline of Australia. It thus becomes a race to find the taxa before they finish their limited fruiting. Some preliminary indications from the Gore Creek flora (Sydney) seem to show that there is a sequence of taxon appearance.

Of intense interest was the Tasmanian component of the Hygrophoraceae which has proven to be 'eye-opening' in a variety of ways. In the Tasmanian forests, the Hygrophoraceae so far documented have almost exclusively been associated with the deep moss beds under *Nothofagus*. This contrasts enormously with the habitats as found so far in Victoria, central New South Wales and southern Queensland where the Hygrophoraceae are found on soil amongst leaf litter. It would be interesting to examine the heath and eucalypt forests in Tasmania under ideal conditions to see if any species do occur.

The most interesting taxon (at least to me) that emerged in troops in most Tasmanian forests was *Hygrocybe astatogala* where it occurred as very large, black basidiomes. Even after encountering this species on numerous occasions, it was still a shock to see it fruit so prolifically on the moss beds. Another very curious occurrence was to find a small quantity of pure white *Hygrophorus involutus*.

An interesting result concerns the two species *Hygrocybe apricosa* and *Hygrocybe aurantiopallens*. These two taxa are very easily separated on the basis of the spores—*Hygrocybe aurantiopallens* has subglobose to globose spores but *Hygrocybe apricosa* has very distinctly ellipsoidal spores. So far, no collection can be assigned to *Hygrocybe apricosa* but there are plentiful collections of *Hygrocybe aurantiopallens* from Tasmania, Victoria and New South Wales. Although there are numerous collections which appear from their colours to be *H. apricosa*, the defining ellipsoidal spores are so far absent from any Australian material.

There remain enormous gaps in the current knowledge of the Hygrophoraceae. Tasmania's flora is still to be thoroughly covered and there may be as many taxa yet again to describe. During 1999, the intention is to revisit Tasmania with the aim of covering only those taxa which are either new or where only one or two collections exist. The next step will be to visit the wet forests of eastern Victoria where little collecting specifically for the Hygrophoraceae has been done and then (if the season permits) visit the Blue Mountains and the Dorrigo area of New South Wales. This year (1998) proved utterly useless as regards collecting in the Dorrigo National Park area as at the time of visiting, the rainforest was so dry that not a single agaric had emerged and only one old, insect riddled polypore was visible on a log.

There also remain enormous gaps just in collection of the species. Almost no collections exist for the northern tropics and there are most definitely species of the Hygrophoraceae to be found in those regions. I have collections and data from northern coastal Queensland and some written data from Darwin which suggests that there remains some very interesting work to be done in the tropics. Western Australia also is likely to prove fascinating. Some data from Katrina Syme and Neale Bougher suggests that there are quite a few undescribed species in the Western Australian flora.

A number of people assisted me during my ABRS work for 1998 and I should very much like to thank them publicly for the enormous support they provided during my collection trip which made my work not only so much easier but in many instances made it actually possible. For the Tasmanian section, I find it very difficult to find words that allow me to thank Alan Mills of the University of Tasmania sufficiently for his exceptionally generous gift to me of his time, his knowledge of the local forests and the use of his facilities, without which the Tasmanian section of my work would have suffered greatly. It would be accurate to say that Alan's assistance virtually doubled the effectiveness of the Tasmanian section. In Victoria, my thanks to Tom May who enthused the participants of the Wilson's Promontory expedition into making sure I got out to Chinaman's Creek and the lush gully at that point that yielded so many taxa within the family. I should also like to pay thankful and immense tribute to the untiring efforts of Ray and Elma Kearney whose dedication to the Hygrophoraceae of Gore Creek in Sydney have not only produced about 20 taxa in a small area of less than half an acre of ground but have contributed at least three new species. My thanks also to Frank Taeker who assisted with collection trips in the Blue Mountains and Royal National Park. And last but not least, Bettye Rees who 'Gymnotonised' her way through Tasmania with Alan and myself and who allowed me to impose very extensively on her hospitality while I was in Sydney. To all of these people again, my sincere thanks.

Reference

Young, A.M. & Wood, A.E. (1977) Studies on the Hygrophoraceae (Fungi, Homobasidiomycetes, Agaricales) of Australia. *Australian Systematic Botany* **10**(6), 911–1030.

MINUTES
AUSTRALASIAN MYCOLOGICAL SOCIETY
COUNCIL MEETING 1997

Held at University of Adelaide, Adelaide, 29 September 1997.

Meeting opened at 5 p.m.

Present: Jack Simpson (chair), Peter Buchanan, David Ellis, Cheryl Grgurinovic, Heino Lepp, Tom May.

1. Minutes of 1996 Council meeting

Accepted.

2. Treasurers Report

Heino Lepp reported that subscriptions totalled approximately \$1800. The four Newsletters each year cost about \$400 each, and with other fixed expenses such as auditing (\$100) the income barely exceeds expenses. Heino recommended an increase in subscriptions. It was also noted that there is an annual drop out rate of members of about 10 per cent, and that there can be quite a delay before subscriptions arrive. Councillors agreed that the concessional rate should be extended to pensioners.

MOTION: Increase subscriptions 50 per cent, and extend concessional rate to pensioners. Moved Heino Lepp, Seconded David Ellis. Carried.

3. Election of Office Bearers

Tom May reported that, although a note in *Australasian Mycological Newsletter* 16(2) p. 41 indicated that current office holders were unable to nominate for their current positions; this was incorrect, and according to the rules of the Society, the President and Vice-President can hold their positions for 3 consecutive years, and other Office bearers and Councillors can remain on Council for up to six consecutive years. Thus the term of the current President and Vice-President expires at the 1998 AGM, although at that time they can nominate for other positions. A single nomination was received for each position, and therefore the following were elected unopposed:

President: Jack Simpson

Vice-President: Cheryl Grgurinovic

Treasurer: Heino Lepp

Secretary: Tom May

Councillors: Peter Buchanan and David Ellis

Ken Thomas is the Public Officer.

4. Special Interest Groups/Subcommittees

(a) Conservation Subcommittee

MOTION: That a Conservation Subcommittee be set up. The membership be the same as the current IUCN, Species Survival Commission, Committee for Fungi, Australasian Regional Committee. The Conservation Subcommittee to continue to report to IUCN. Moved Jack Simpson, Seconded Cheryl Grgurinovic. Carried.

(b) There is a need to establish a further subcommittee to raise the profile of and promote mycology. Issues to be discussed include ways of funding mycology, and the prospects for an Australian National Institute of Fungal Biodiversity. TM to seek members of this subcommittee.

5. Meetings

1998. No scientific meeting is planned. The NZ fungus foray is in the central North Island, and difficult to get to. Seems best to just have an AGM/Council meeting in 1998.

1999. The scientific meeting will be held in conjunction with the IUMS Congress of Mycology, Sydney, August 16–20. There will be a plenary session in the IUMS Congress on biodiversity and biogeography of Australasian fungi, and a number of symposia. There should be a day for students and other AMS members to present papers and posters.

2000. Could have a scientific meeting joint with the NZ Foray.

AGM must be held once a year, and according to the constitution must be held between July and November inclusive. In a year where there is no Scientific meeting (as for 1998), or it is held in the first half of the year (as may be the case in 2000), the AGM can be held at some place and time convenient to council members. When there is a Scientific meeting without an AGM, a General Meeting can be held at the time of the Scientific Meeting, to enable members to discuss issues. The quorum for an AGM is five, including at least two Council members. The quorum for Council is four, or else three with at least one proxy.

200th anniversary of systematic mycology in Australia is in 2000 (*Aseroe rubra* described in 1800). TM suggested that a publication celebrating this could include a survey of the state of systematic mycology in the region, covering all major groups of the mycota. Year 2000 (Federation of Australia anniversary) projects are a possible source of funding. This publication could result from the 1999 conference, or be additional or separate.

IMC 2006. IMC 2002 is in Oslo. Eric McKenzie chairs a committee putting together an Australasian bid for IMC 2006. Support is sought from APPS, ASM. Should also approach lichenologists. In regard to conference organisation—John Pitt is enthusiastic about the organisers of IUMS Sydney Congress. Tour hosts are not favoured by David Ellis, due to problems with contracts and budget. There should be the ability to confirm receipts. A decision on how to arrange the IMC must lie with the Conference Organising Committee. It will be necessary to have an Australasian city chosen before the Jerusalem IMC. Auckland is not a possibility due to lack of accommodation. The best time seems to be September, because this is traditional. An autumn date remains a possibility.

6. Patron

It was agreed that the Society should have a patron, and that Jack Warcup would be an excellent choice.

MOTION: That Jack Warcup be made patron of the Australasian Mycological Society. Moved Jack Simpson, Seconded David Ellis. Carried.

7. Web Page

CG happy to set up web page. ABRS staff will be having training on web pages, and ABRS happy for web page to sit on ABRS server.

MOTION: Cheryl Grgurinovic to set up Australasian Mycological Society web page. Moved David Ellis, Seconded Heino Lepp. Carried.

8. Payments by NZ members

Suggested options are:

(1) pay into Landcare or Foray account, then (a) PB to send on to AMS account, or (b) retain in NZ, and then use for direct support of Society activities in NZ, with pro forma receipt available for auditing. Would need to check if (b) was acceptable from an accounting point of view.

(2) set up a NZ subaccount. Option (2) would only be viable with a reasonable sum of money due to bank fees.

NZ exchange rate is currently about 0.87.

MOTION: New Zealand members of the Australasian Mycological Society send subscriptions to Peter Buchanan at the same amount in \$NZ as the \$AUD subscription rate. Peter to send the bulk subscriptions to the Treasurer before the end of each financial year. Moved Jack Simpson, Seconded Peter Buchanan. Carried.

9. Renewal form

It was agreed that it would be useful to have the amount of the subscription shown on the renewal form.

10. Sustaining Members and Advertising

DE gave the example of the American Mycological Society and the Australian Society of Microbiology, where sustaining members pay \$100–\$400 per annum. The introduction of sustaining members may necessitate alterations to the Constitution. Should look out for commercial opportunities, such as advertising in the *Newsletter*, or inclusion of book fliers. Items that are educational, or relate to publications by financial members to remain free of charge.

MOTION: That the Society allow sustaining members, with an annual subscription of \$100. Moved David Ellis, Seconded Cheryl Grgurinovic. Carried.

11. IMA Asian Group

There has been no response from IMA Asian group to enquiries from AMS.

12. Local chapters

Should encourage meetings in major centres.

Meeting closed 7.15 p.m.

T.W. May



AUSTRALASIAN MYCOLOGICAL SOCIETY INC.

ANNUAL GENERAL MEETING

13 NOVEMBER 1998

MINUTES

Held at: Lecture Room 4th Floor, School of Biological Sciences, University of New South Wales.

Meeting opened 6.30 p.m.

Present: Jack Simpson (chair), Cheryl Grgurinovic, Tom May, Wieland Meyer, and 17 other members and friends.

1. Minutes of 1997 AGM

MOTION: Minutes of 1997 AGM accepted as true and accurate record. Moved: Peter McGee, Seconded: Cheryl Grgurinovic. Carried.

2. New office bearers

Since a single nomination has been received for each position, the following will take office:

President: Cheryl Grgurinovic, Vice-President: Wieland Meyer, Treasurer: Heino Lepp, Secretary: Tom May, Councillors: Peter Buchanan, Ceri Pearce.

A welcome was extended to the two new Council members: Ceri Pearce, working at CSIRO Atherton on a revision of *Phyllachora*, and Wieland Meyer, working at Westmead Hospital Sydney, on molecular taxonomy of fungi, particularly yeasts such as *Candida*. The current Council of AMS now comprises a good balance in terms of taxa (macrofungi, microfungi) and interests (taxonomy, ecology, conservation, quarantine, plant pathology, and medical mycology).

3. Financial report

The financial report for the year ended 30 June 1998 was tabled, showing a surplus of \$1544 for the year ended 30 June 1998, and total assets of \$6780. MOTION: that the Financial report be accepted. Moved: Anne Ashford, Seconded: Wieland Meyer. Carried.

4. AMS activities

4a. Publications

Jack Simpson reported that the *Australasian Mycological Newsletter* continues to be published quarterly. It is proposed to alter the name of the Society publication to *Australasian Mycologist* from the first issue of 1999, and to encourage a greater proportion of refereed contributions.

4b. Web site

Cheryl Grgurinovic has prepared a web site for the Society which will be accessible shortly.

4c. International Mycological Society

Jack Simpson reported that AMS is an affiliated Society of the International Mycological Association, which does not give AMS voting rights. Full membership of IMA is \$US400 per year, and considered too large an outlay for AMS at present.

5. Future meetings

5a. 1999—Sydney

A scientific meeting of AMS will be organised in association with the IUMS Congress of Mycology which is being held in Sydney in August 1999. A likely date is Saturday 21 August. This will provide an inexpensive opportunity for students and others not presenting papers at the IUMS conference to talk about their work to mycological colleagues. Note that there are a limited number of vacancies in the IUMS program for speakers, so members are encouraged to submit proposals to Symposium convenors (see AMS 17 for full list of symposia), although most will have to be presented as posters. There will be a foray to Watagan Forest on the Saturday before the IUMS conference (August 14), organised by Jack Simpson, with participation from the Sydney Fungal Studies Group.

5b. 2000—Te Anau

The scientific meeting of AMS in 2000 will be held in association with the New Zealand fungal foray, planned for 30 April–6 May, at Lake Te Anau, Fiordland. The Foray organiser is Lawrie Taylor. During the foray, AMS will hold a one day scientific meeting and an AGM.

5c. 2006—IMC

IMC7 is in Oslo in 2002. There is a strong possibility that IMC8 (due to be held in 2006) and the IUMS Mycology Congress will be held jointly. Due to the IUMS Mycology Congress already having been held in Australia in 1999, a bid for any such 'joint' mycology congress is unlikely to be successful. In case a joint congress does not eventuate, an Australasian bid for IMC8 is 'in the wings'.

6. Other business**6a. Vote of thanks to outgoing Councillors**

Cheryl Grgurinovic took the chair as incoming President and proposed a vote of thanks to Jack Simpson and David Ellis, who are retiring from Council. Tom May praised the vision of Jack Simpson in playing a large role in setting up the Society and ensuring its continuation.

Meeting closed at 7.00 p.m.

SIGNED (Jack Simpson)

REPORTS OF SPECIAL INTEREST GROUPS

Conservation Subcommittee. Tom May has taken up the role of coordinator of this group. The *Conservation Overview of Australian non-marine Lichens, Bryophytes, Algae and Fungi*, published in 1997, is currently being examined by Environment Australia to see how best to implement the recommendations. The Subcommittee will make a submission on suggested actions to improve the conservation status of Australian fungi.

Education Special Interest Group. Peter McGee reports that the Group submitted an application to CUTSD for support for initial development of a national education program to be delivered over the WWW. The application was forwarded through the University of Sydney and supported by them. Unfortunately, the application was unsuccessful. The Group is now investigating alternative approaches to raising funds, for two reasons. The funds for CUTSD are limited, and would not have allowed completion of the program. Secondly, the program must be completed within 12 months, making it difficult to continue constructing the package, and refining the materials.

Fungal Poisoning Network. Tony Young continues to act as Convenor of the Poisoning Network. He reports that some issues relating to liability insurance for those offering identifications and information in regard to fungal poisonings remain unresolved. Individuals may need their own insurance, where they are not associated with an institution. Even when there is institutional affiliation, out of state inquiries may not strictly be covered by liability insurance.

Tom May
Secretary, Australasian Mycological Society

AUSTRALASIAN MYCOLOGICAL SOCIETY

ACCOUNTS

THE 13th NEW ZEALAND FUNGAL FORAY

**Forrestal Lodge, Inglewood, Mt Egmont National Park,
evening of Wednesday 28 April to morning of Sunday 2 May 1999**

Mt Taranaki (Egmont) is a dormant volcano which last erupted in 1655. With an altitude of 2518 m it is high enough to have a wide range of climates, from mild lowland to severe alpine, with a vegetation sequence from lowland forest to alpine herbfields. Rainfall is high ranging from 1150 mm in the surrounding plain to 6500 mm at Dawson Falls on the mountain. At the park boundary, at 550 m, the broadleaf/ podocarp forest is dominated by kamahi (*Weinmannia racemosa*), but also present are tawa (*Beilschmiedia tawa*), miro (*Prumnopitys ferruginea*), hinau (*Elaeocarpus dentatus*), rimu (*Dacrydium cupressinum*) and rata (*Metrosideros robusta*). The upland forest continues to be dominated by kamahi but with kapuka (*Griselinia littoralis*), kotukutuku (*Fuchsia excorticata*) and orihou (*Pseudopanax colensoi*). At 900 m kamahi begins to be replaced by Halls totara (*Podocarpus halli*) and kaikawaka (*Libocedrus bidwilli*). These in turn are replaced by alpine shrublands, tussock herbfields and then mossy herbfields. Mycologically the area has only been lightly explored.

The Foray will stay at Forrestal Lodge located at 23 Rimu St in Inglewood township in a former convent. It is carpeted, heated, and the rooms are two and three bedded.

The cost of accommoation will be about \$12 per night, plus the additional cost for food (all meals supplied).

A limited number of grants towards the daily cost and/or transport to the foray are available for students. More information is available from Geoff Ridley.

Geoff Ridley, NZ Forest Research Institute, Private Bag 3020, Rotorua, New Zealand
tel: +64 7 347 5899; fax: +64 7 347 5333; e-mail: ridleyg@fri.cri.nz

Registration for the 13th New Zealand Fungal Foray, Mt Egmont National Park, 28 April—2 May 1999

Name:.....

Address:.....

.....

.....

Tel:.....

Fax:.....

E-mail:.....

Number attending:.....

I will be attending the full foray and require accommodation and meals Yes/No

I will be a day visitor and require meals only Yes/No

I will be a day visitor and do not require any meals Yes/No

Deposit (\$40/person) enclosed: ***NZ dollars***

PLEASE MAKE CHEQUES PAYABLE TO 'FORAY ACCOUNT'

PLEASE FORWARD THIS MESSAGE TO ANYONE YOU THINK MAY BE INTERESTED

NEW MEMBERS**Full members:**

Dr Brett Summerell, Royal Botanic Gardens, Sydney
 Ms Sook Hahn Soo, Gosford, NSW
 Dr Egon Horak, Zurich
 Jim Gardner, Parramatta, NSW

NEW TITLE FOR AUSTRALASIAN MYCOLOGICAL NEWSLETTER

From the March issue in 1999 (issue 18(1)) the *Australasian Mycological Newsletter* will change its name to the *Australasian Mycologist*. Volumation will continue without change.

MEMBERSHIP DIRECTORY

The editors would like to publish a separate membership directory this year. If you **do not** want your name and address to be included in the membership directory please contact Cheryl Grgurinovic at the address on the verso of the front cover. For the membership directory to be as up to date as possible, if you do wish to appear in the directory please make sure you include your current address, phone and fax numbers and email address on the membership renewal form.

AUSTRALASIAN MYCOLOGICAL SOCIETY MEETING

**to be held in conjunction with the International Union of Microbiological Societies,
 IXth International Congress of Mycology**

The Australasian Mycological Society will be holding a one day meeting on Saturday 21 August 1999 at Westmead Hospital. Further details and the registration form will be in the March issue of the *Australasian Mycologist*.

CONFERENCES AND WORKSHOPS

9–12 February 1999	Grand Mercure Hotel, Gold Coast, Qld	First Australasian Soilborne Disease Symposium	ASDS Secretariat PO Box 717 Indooroopilly, Qld 4068 Ph.: 07 3878 9242 Fax: 07 3878 9530 Email: yrdpco@ozemail.com.au
22–23 February 1999	CABI Bioscience	Basic Mycological Techniques	Mrs Stephanie Groundwater, CABI Bioscience UK Centre, Egham, Surrey, TW20 9TY, UK Ph.: +44 (0) 1784 470111 Fax: +44 (0) 1491 829100 Email: S.Groundwater@CABI.org (Please give your postal address.)
1–5 March 1999	CABI Bioscience	Identification of Industrial and Food Spoilage Fungi	Mrs Stephanie Groundwater, CABI (Address given above.)
4–6 March 1999	Aix-en, France	Fifth International Congress for the Science and Cultivation of <i>Tuber</i> and other edible hypogeous mushrooms	courvoisier@wanadoo.fr for registration information
16–18 March 1999	CABI Bioscience	Fungi Associated with Water Systems	Mrs Stephanie Groundwater, CABI (Address given above.)

23–28 March 1999	Asilomar Conference Center, California	20th Fungal Genetics Conference	Kevin McCluskey VOX 913.588.7044 FAX 913.588.7295 fgsc@kuhub.cc.ukans.edu http://www.kumc.edu/research/fgsc/asilo99/announc.html
28 June–9 July 1999	CABI Bioscience	Biochemical and Molecular Characterisation of Bacteria and Fungi	Mrs Stephanie Groundwater, CABI (Address given above.)
12 July–13 August 1999	CABI Bioscience	International Course on the Identification of Fungi of Agricultural and Environmental Significance	Mrs Stephanie Groundwater, CABI (Address given above.)
26–30 July 1999	Beltsville, Maryland, USA	The Third International Congress on the Systematics and Ecology of Myxomycetes	Lafayette Frederick Biology Department Howard University Washington, DC 20059 or Steve Stephenson Department of Biology Fairmont State College Fairmont, WV 26554, USA <sls@fscvax.wvnet.edu>
1–7 August 1999	St Louis, MO, USA	XVI International Botanical Congress	http://www.abc99.org/
16–20 August 1999	Sydney	IUMS IXth International Congress of Mycology	IUMS Secretariat GPO Box 128 Sydney NSW Australia 2001 email: iums@tourhosts.com.au ph: 61 2 9248 0812 fax: 61 2 9262 3135 ixicbamm99@tourhosts.com.au http://biology.anu.edu.au/iums/
31 August–3 September 1999	Caracas, Venezuela	III Congreso de la Asociación Latino- Americana de Micología	Calle Garcilazo, c/o Chama, Centrol Polo Torre A, Piso 8, Ofc 83, Caracas 1050, Venezuela VOX/FAX +58.2.751.8605/8338/5629 event@eldish.net
21–25 September 1999	University of Alcalá, Madrid, Spain	The XIII Congress of European Mycologists	http://www.cicom.es/fundacion/micolog.htm
27 September–1 October 1999	Canberra	APPS 12th Biennial Conference	Philippa Rowland Conference Secretary Ph. 6272 3443; Fax: 6272 4896; email < pcr@brs.gov.au >
11–16 October 1999	Sydney	3rd International Conference on Mushroom Biology and Mushroom Products	Conference Secretariat: 405 Bull Ridge Road East Kurrajong, NSW 2758 FAX +61 2 4576 3610 bkgregg@zeta.org.au

14–20 November 1999	Yaounde, Cameroon	2nd International Virology and Microbiology Conference	Dr Njayou Mounjohou B.P. 2001 Messa, Yaounde VOX 237.21.43.10 FAX 237.23.37.09 ebola@camnet.cm
6–10 December 1999	Perth, WA	Society of Australian Systematic Biologists, Australian Systematic Botany Society, Invertebrate Biodiversity and Conservation 1999 joint conference: 300 years in New Holland and Old Australia	—
25–29 March 2000	John Moore's University, Liverpool, UK	BMS Millennium Symposium on Tropical Mycology	—
9–14 July 2000	University of Hong Kong	2nd Asia-Pacific Mycological Conference on Biodiversity and Biotechnology	Kevin Hyde Email: kdhyde@hkucc.hku.hk
7–12 August 2000	Kuala Lumpur, Malaysia	XXI IUFRO World Congress Forests and Society: the role of research	Chairman The XXI IUFRO World Congress Organising Committee Forest Research Institute Malaysia (FRIM) Kepong, 52109 Kuala Lumpur Malaysia Fax: 603 6365687/6367753 iufroxxi.csc@forvie.ac.at http://iufro.boku.ac.at/iufro/congress/csc/
August 2002	University of Oslo, Norway	7th International Mycological Congress	Leif Ryvarden Botany Department Biological Institute Box 1045 Blindern, N-0316 Norway Ph.: 47-22854623 Fax: 47-22856717 leif.ryvarden@bio.uio.no

If you know of any other conferences, symposia, workshops, *etc.* that may be of interest to members, please send us the details so the information can be included in the next *Newsletter*.

Cheryl Grgurinovic

MYCOSURFING ON THE WORLD WIDE WEB

Australasian Mycological Society website

This website will be available soon. I would like to include a page of recent publications by members (since January 1998). Please write to or email me at the address on the verso of the front page if you would like to have your recent publications included on the website.

C. Grgurinovic

DEADLINE FOR NEXT ISSUE

Articles for the next *Newsletter* are due by Friday 12 March 1999. If articles are more than half a page long, the editors would appreciate a copy on disc. **Please note that *journal and book titles are given in full in the references.* The editors request that authors adhere to the *Newsletter* style of citing references.**

Manuscripts submitted for publication in the *Australasian Mycological Newsletter* should not be submitted to another journal waiting publication or have been previously published in another journal.

RENEWING MEMBERS OF THE AUSTRALASIAN MYCOLOGICAL SOCIETY, INC.

Membership subscriptions are due on 1 January 1999. If you have not renewed your suscription by March 1999, this will be your last copy of the *Newsletter*.

Membership subscription in the Society for 1999 is AUS\$30 per calendar year for Full Members and AUS\$15 for Student Members and Retired Members in Australia or New Zealand; AUS\$45 for Full Members and AUS\$30 for Student Members and Retired Members outside Australia or New Zealand. Subscriptions include four issues per year of the *Australasian Mycological Newsletter* and postage charges. Subscriptions fall due on 1 January of each year.

Library subscription AUS\$45 per calendar year; personal members are requested not to donate their copies of the *Newsletter* to a library for 12 months from publication date.

Subscriptions

Other than New Zealand members:

Treasurer
Mr Heino Lepp
PO Box 38
BELCONNEN, ACT 2616, Australia.
*Please make cheques payable to the
Australasian Mycological Society, Inc.*

Subscriptions

New Zealand members:

New Zealand members can pay subscriptions in \$NZ. Send cheques to
Dr Peter Buchanan at Landcare Research, Private Bag 92170, Auckland,
New Zealand.
Please make cheques payable to 'Foray account'.

✂ _____

Name:.....

Address:.....

.....

.....

.....

Phone Number:.....

Fax number:.....

Email address:.....

✂ _____

NEW MEMBERS OF THE AUSTRALASIAN MYCOLOGICAL SOCIETY, INC.

AUSTRALASIAN MYCOLOGICAL SOCIETY INCORPORATED

(incorporated under the Associations Incorporation Act 1991)

APPLICATION FOR MEMBERSHIP

I,

.....

of

.....

(address)

Phone number:.....

Fax number:.....

Email address:.....

.....hereby apply
to

(occupation)

(full time students must show evidence of enrollment at a secondary or tertiary institution)

become a member of the abovenamed incorporated association. In the event of my admission as a member, I agree to be bound by the rules of the Society for the time being in force.

.....

(signature of applicant)

Date

I,

(full name)

a member of the Society, nominate the applicant, who is personally known to me, for membership of the Society.

.....

(signature of proposer)

Date

I,

(full name)

a member of the Society, nominate the applicant, who is personally known to me, for membership of the Society.

.....

(signature of proposer)

Date

Subscriptions

Other than New Zealand members:

Treasurer

Mr Heino Lepp

PO Box 38

BELCONNEN, ACT 2616, Australia. *Please make cheques payable to the Australasian Mycological Society, Inc.*

Subscriptions

New Zealand members:

New Zealand members can pay subscriptions in

\$NZ. Send cheques to Dr Peter Buchanan at

Landcare Research, Private Bag 92170, Auckland,

New Zealand.

Please make cheques payable to 'Foray account'.

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